



# Microbial groups

CE 421/521

Lecture September 14, 2006

Vaccari et al., Chapter 10



# Microbes

- Microorganisms – broad category of organisms too small to be seen with the naked eye
- Integral part of every ecosystem
- Roughly  $10^6$  to  $10^9$  per gram of soil, biofilm or sludge sample



# Microbial groups

## ■ Prokaryotes

- Bacteria (Including blue green algae)
- Archae – (sometimes archaeobacteria)
  - classified during the 1970's by Carl Woese and George Fox
  - Don't fit neatly into prokaryotic or eukaryotic class due to their difference in 16S rRNA - separate kingdom?
  - Includes methanogens and halophiles

■ Viruses – Dimitri Ivanovsky (1893) filtered sap through ceramic filters designed to remove bacteria – still resulted in tobacco mosaic virus

## ■ Eukaryotes



# Classification of microorganisms

## ■ Energy source:

- Chemotrophs – energy from chemical substances
  - Organotrophs – energy from organic compounds
  - Lithotrophs – energy from an inorganic compound
- Phototrophs – energy from sunlight



# Classification of microorganisms

## ■ Carbon source:

- Heterotrophs – carbon from organic compounds
- Autotrophs – carbon from inorganic compounds

Can have mixed classifications:

e.g. chemoorganoheterotroph (example E. Coli)

chemolithoautotroph (example nitrobacter)



# Classification of microorganisms

## ■ Environmental preferences:

- TEA (anaerobic, aerobic, anoxic)

- Temperature

- Psychrophiles
- Mesophiles
- Thermophiles

- pH

- Neutrophiles (5-9)
- Acidophiles ( $< 5$ )
- Alkaliphiles ( $> 9$ )

## ■ Extremophiles – can grow at extreme temperatures or osmotic pressures (e.g., halophiles)



# Microbial Taxonomy

- Morphology: form and visible structure
  - Biochemical activities
  - Phenotype – representing observable characteristics
- Genotype
  - Characterized by DNA or RNA
  - Phylogeny – based on genetic similarities




# Taxonomy - What is a prokaryotic species?

- Difficulty in that genetic exchange occurs between species not necessarily closely related
  - Strain
    - have a recent parent cell
    - Share genetic properties with minor exceptions
  - Species
    - Share at least 70% of DNA homology – similarity in DNA sequence
    - Or have rRNA similarity of 97% or greater
  - Genus
    - Share at least 20% of their DNA homology
    - Or have rRNA similarity of 93-95%





# Nomenclature

1. E\_\_\_\_\_ (e.g., *aquaticus*, *marina*, *coli*)
  2. H\_\_\_\_\_ (e.g., *bovus*, *avium*)
  3. Environmental c\_\_\_\_\_ (e.g., *thermophilus*, *halophilus*)
  4. S\_\_\_\_\_ (e.g., *ovalis*, *longum*, *sphaericus*)
  5. C\_\_\_\_\_ (e.g., *aureus*, *niger*)
  6. S\_\_\_\_\_ (e.g., *denitrificans*, *avium*)
  7. P\_\_\_\_\_ (e.g., *methanobacterium*, *cerevisiae*)
  8. D\_\_\_\_\_ (e.g., *typhi*, *botulinum*, *pneumoniae*)
  9. P\_\_\_\_\_ (e.g., *winogradskii*, *burkholderia*)
- 



# Prokaryotes - shape

- cocci (spherical, e.g., *Streptococcus*)
- bacilli (rod shapes, e.g., *Bacillus subtilis*)
- spirilla (spiral, e.g., *Spirillum volutans*)
- filamentous



# Prokaryotes - shape

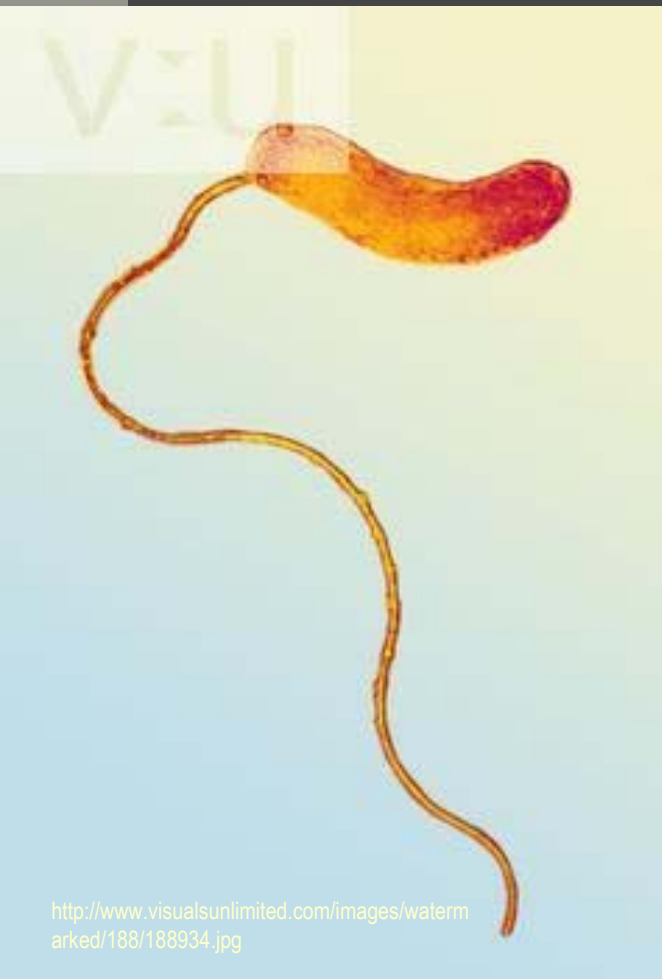
## ■ Unusual

- s\_\_\_\_\_ bacteria - filamentous, surrounded by a sheath
- s\_\_\_\_\_ bacteria - aerobic, gram negative, at end of stalk is a “holdfast” allows it to attach to surfaces
- b\_\_\_\_\_ bacteria, multiply by budding, bud grows flagellum, settles on new surface and buds again
- g\_\_\_\_\_ bacteria, filamentous, gram-negative, “glide” along solids surfaces, *Beggiatoa* and *Thiothrix*: oxidize  $\text{H}_2\text{S}$  to  $\text{S}^0$

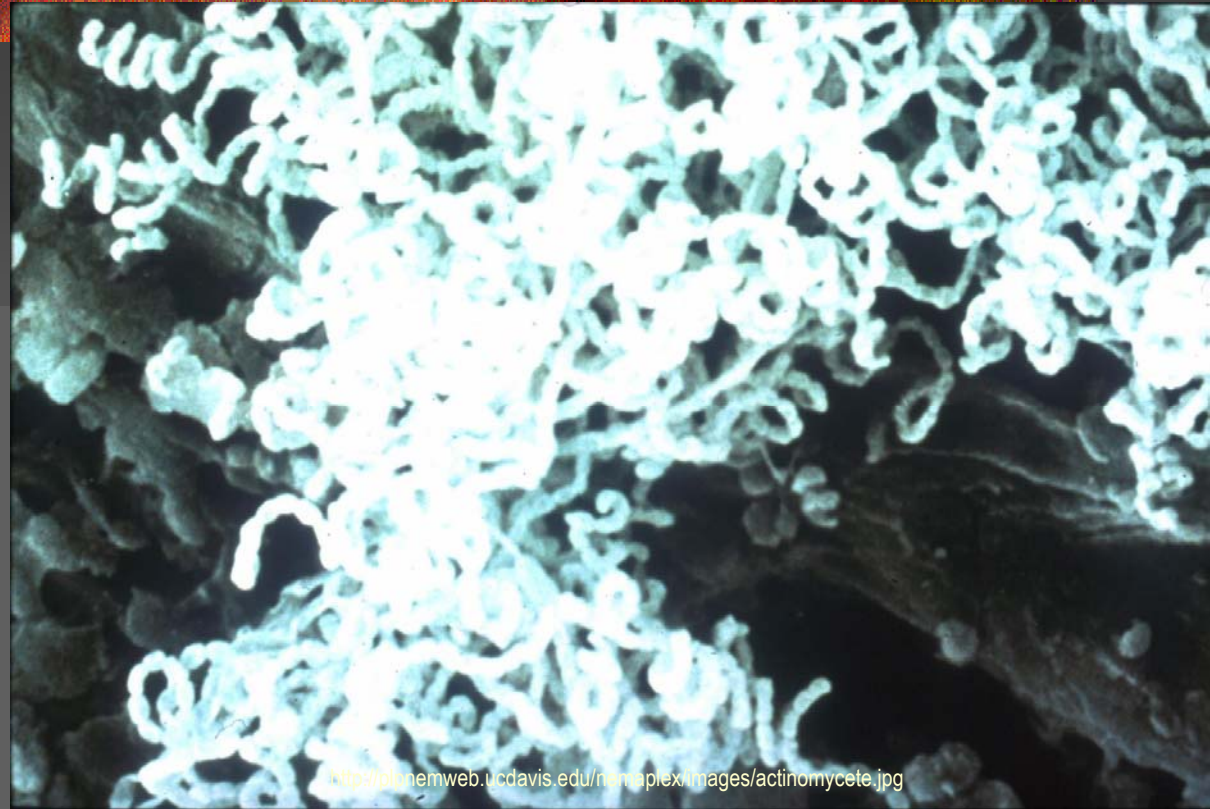


# Prokaryotes - shape

- *Bdellovibrio* - s\_\_\_\_\_ (0.2-0.3 $\mu$ ) flagellated bacteria that prey on gram-negative bacteria
- Actinomycetes- gram-positive, f\_\_\_\_\_, have branching filaments similar to fungi - *Streptomyces* and *Nocardia*
- Cyanobacteria - b\_\_\_\_\_ -g\_\_\_\_\_ algae, procaryotes, contain chlorophyl a, have characteristic blue-green color, contain gas vacuoles that enable them to float to maximize photosynthesis, responsible for algal blooms, some are toxic

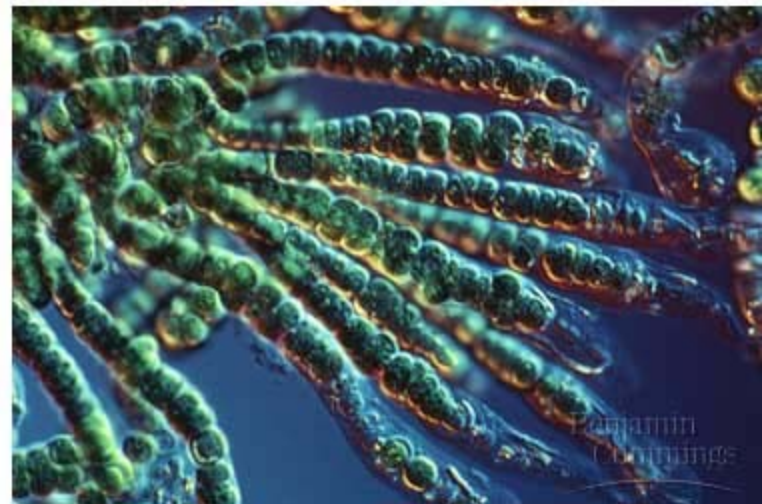
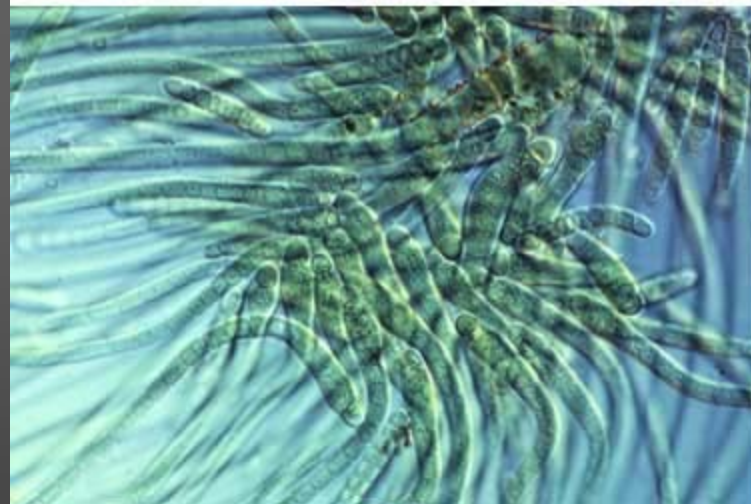


*Bdellovibrio*



Actinomycetes





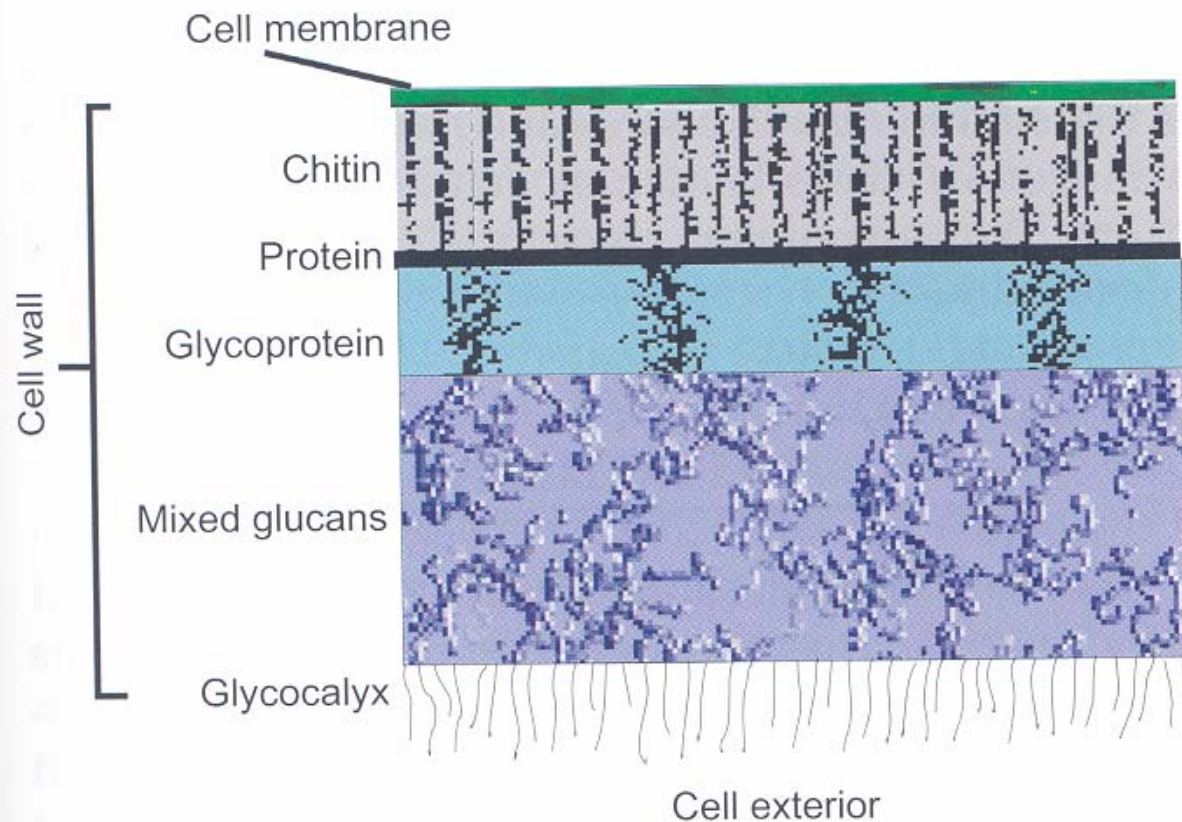
Cyanobacteria



# Fungi

- e\_\_\_\_\_, produce long filaments called hyphae containing c\_\_\_\_\_
- heterotrophs, use o\_\_\_\_\_ compounds for carbon and energy
- found during n\_\_\_\_\_ limitations, low D.O., low pH conditions
- important in the cycling of organics – degradation of plant polymers cellulose and lignin
- primarily aerobic (except for fermentative yeast)

# Fungal cell wall



**FIGURE 2.17** Structure of a fungal cell wall.





# Algae

- most are u\_\_\_\_\_, floating, phytoplankton
- some are f\_\_\_\_\_
- most are p\_\_\_\_\_
- all contain chl\_\_\_\_\_ a, some b and c
- found in o\_\_\_\_\_ ponds, polishing ponds, aerobic lagoons

# Algal Cell

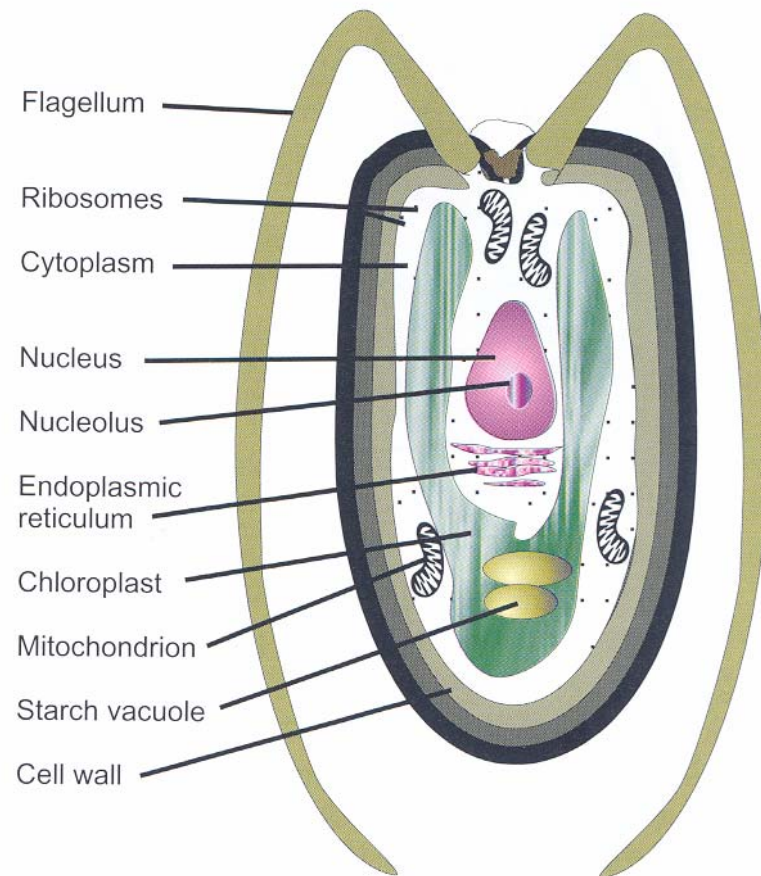
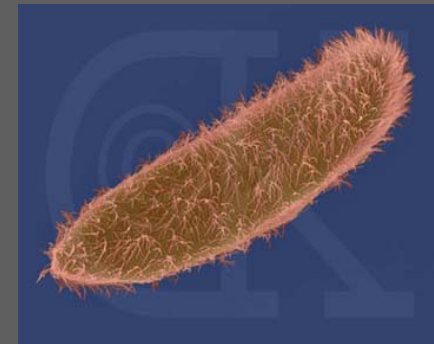
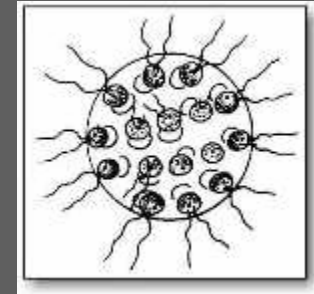
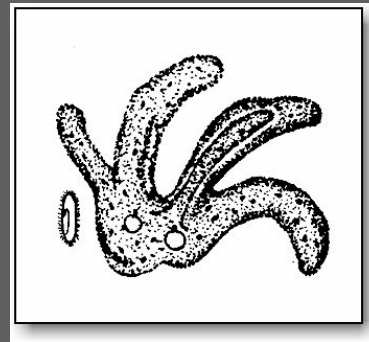


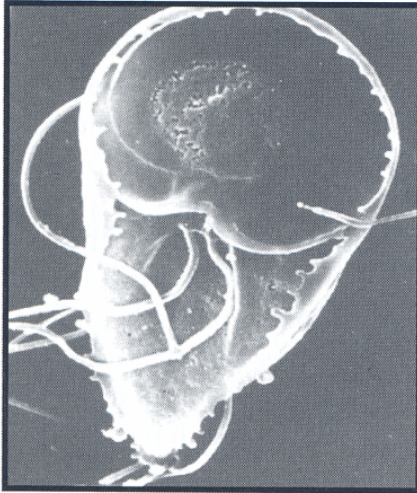
FIGURE 2.18 Structure of a typical algal cell.

# Protozoa

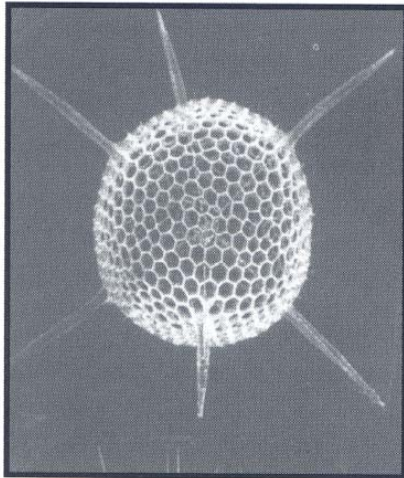
- Unicellular
- Heterotrophs
- Classification
  - sarcodina (amoebae)
  - mastigophora (flagellates)
  - ciliophora (ciliates)
  - sporozoa



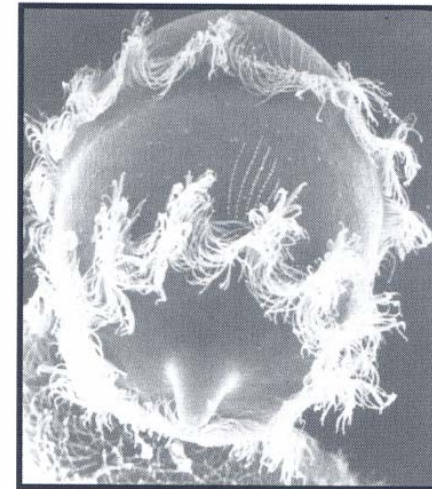
A



B



C



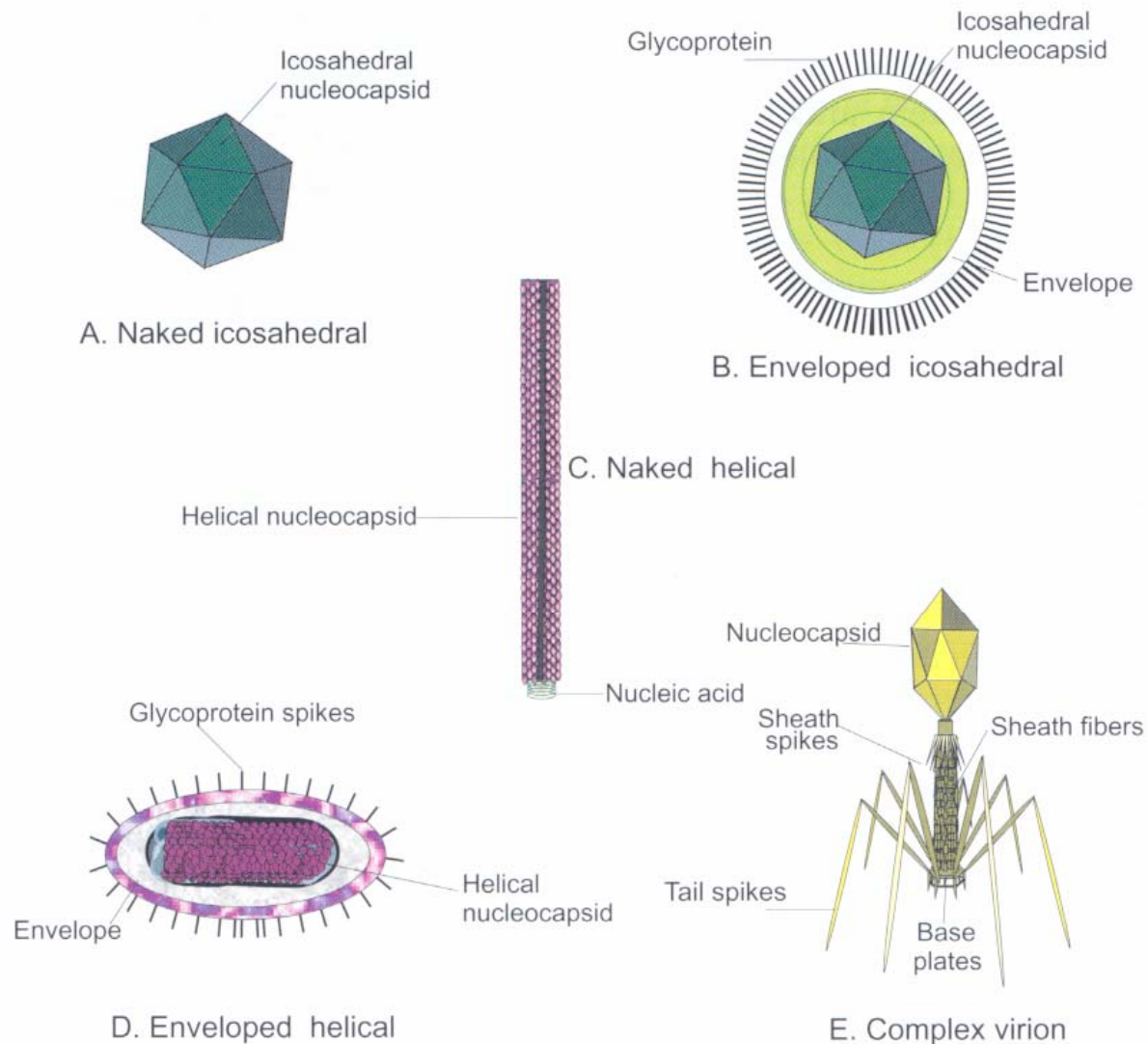
**FIGURE 2.19** Basic morphology of protozoa. (A) scanning electron micrograph of a flagellar protozoa, *Giardia*; (B) scanning electron micrograph of testate cilia, *Heliosoma*; (C) electron micrograph of ciliated *Didinium*. Part A reprinted with permission from Cox (1993). Parts B and C reprinted with permission from Sleight (1989).



# Viruses

- small c\_\_\_\_\_ particles (not procaryotes or eucaryotes) are they alive?
- replication occurs in h\_\_\_\_\_
- Structure
  - c\_\_\_\_\_ of nucleic acid (could be double or single stranded, DNA or RNA) surrounded by protein coat (capsid)
  - main shapes
    - h\_\_\_\_\_
    - p\_\_\_\_\_
    - c\_\_\_\_\_





**FIGURE 2.4** Simple forms of viruses and their components. The naked icosahedral viruses (A) resemble small crystals: the enveloped icosahedral viruses (B) are made up of icosahedral nucleocapsids surrounded by the envelope: naked helical viruses (C) resemble rods with a fine regular helical pattern in their surface: enveloped helical viruses (D) are helical nucleocapsids surrounded by the envelope: and complex viruses (E) are mixtures of helical and icosahedral and other structural shapes.



# Virus Replication

1. Ad\_\_\_\_\_ - virus adsorbs to specific receptors, receptors can be polysaccharides, proteins, or lipoproteins
2. En\_\_\_\_\_ - various particle or nucleic acid material enters cell
3. Ec\_\_\_\_\_ - capsid is stripped away, releasing genetic material
4. Mu\_\_\_\_\_ - viral nucleic acids are replicated using machinery of host cell
5. Ma\_\_\_\_\_ - protein coat is synthesized and combined with nucleic acid to form nucleocapsid
6. Re\_\_\_\_\_ of mature virions - host cell ruptures release active viruses



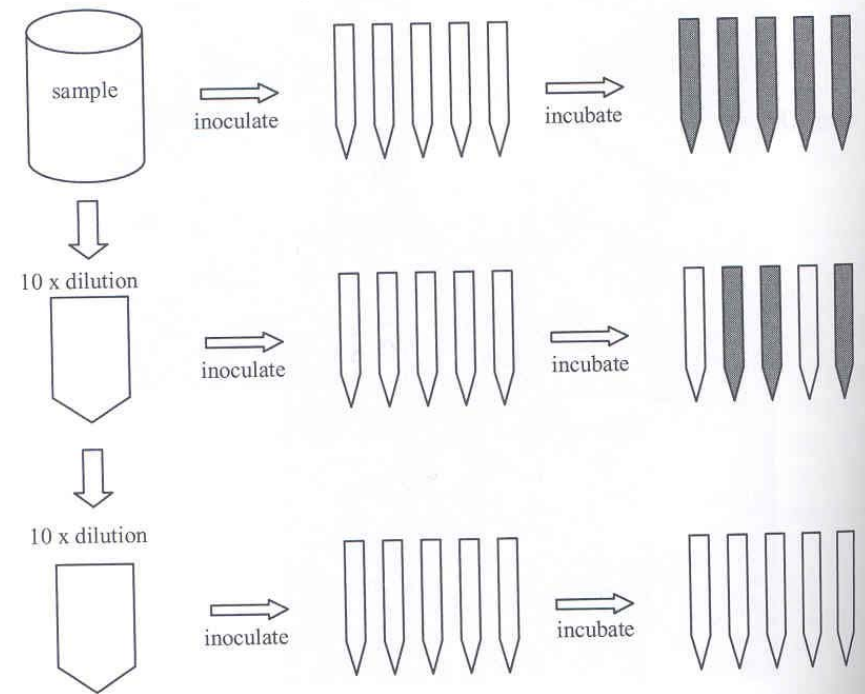
# Virus Detection and Enumeration

- animal i\_\_\_\_\_ - newborn mice injected with inoculum and observed for signs of disease
- t\_\_\_\_\_ cultures - viruses quantified by measuring effect on host cell lines forming a monolayer on glass or plastic assay bottles, effect is measure by
- p\_\_\_\_\_ assay - virus is placed on surface of host cell monolayer, virus replication leads to localized area of cell destruction called plaques
- s\_\_\_\_\_ dilution endpoint - virus suspension is diluted serially and the highest dilution (smallest amount of virus) that causes a cytopathic effect in 50% of samples is reported as the tissue culture infectious dose (TCID50)
- most p\_\_\_\_\_ number - serial dilutions placed in tubes or microwells with host cells, positive tubes are recorded and MPN value computed from standardized MPN table.



# MPN

- Uses serial dilutions and statistical probabilities for the most likely number of organisms giving a positive response



**Figure 11.13** MPN method schematic. Positive tubes are shown as dark after incubation.

Example: Take 1 mL of sample and add to 1 L of water then perform the following serial dilutions: 10 mL, 1 mL, and 0.1 mL and incubate with substrate. If we get 5 positive tubes in the first dilution, 4 positive tubes in the second dilution, and 1 in the last dilution, what is the MPN of the sample?

Solution: from the following table we can see that the as diluted MPN is 170. Since we had a 1000 fold dilution to start with, the resulting MPN is 170,000 organisms per 100 mL

Number of positive tubes				Number of positive tubes				Number of positive tubes				Number of positive tubes				Number of positive tubes				Number of positive tubes			
10 mL	1 mL	0.1 mL	MPN	10 mL	1 mL	0.1 mL	MPN	10 mL	1 mL	0.1 mL	MPN	10 mL	1 mL	0.1 mL	MPN	10 mL	1 mL	0.1 mL	MPN	10 mL	1 mL	0.1 mL	MPN
0	3	0	5.6	1	3	0	8.3	2	3	0	12	3	3	0	17	4	3	0	27	5	3	0	79
0	3	1	7.4	1	3	1	10	2	3	1	14	3	3	1	21	4	3	1	33	5	3	1	110
0	3	2	9.3	1	3	2	13	2	3	2	17	3	3	2	24	4	3	2	39	5	3	2	140
0	3	3	11	1	3	3	15	2	3	3	20	3	3	3	28	4	3	3	45	5	3	3	180
0	3	4	13	1	3	4	17	2	3	4	22	3	3	4	31	4	3	4	52	5	3	4	210
0	3	5	15	1	3	5	19	2	3	5	25	3	3	5	35	4	3	5	59	5	3	5	250
0	4	0	7.5	1	4	0	11	2	4	0	15	3	4	0	21	4	4	0	34	5	4	0	130
0	4	1	9.4	1	4	1	13	2	4	1	17	3	4	1	24	4	4	1	40	5	4	1	170
0	4	2	11	1	4	2	15	2	4	2	20	3	4	2	28	4	4	2	47	5	4	2	220
0	4	3	13	1	4	3	17	2	4	3	23	3	4	3	32	4	4	3	54	5	4	3	280
0	4	4	15	1	4	4	19	2	4	4	25	3	4	4	36	4	4	4	62	5	4	4	350
0	4	5	17	1	4	5	22	2	4	5	28	3	4	5	40	4	4	5	69	5	4	5	430
0	5	0	9.4	1	5	0	13	2	5	0	17	3	5	0	25	4	5	0	41	5	5	0	240
0	5	1	11	1	5	1	15	2	5	1	20	3	5	1	29	4	5	1	48	5	5	1	350
0	5	2	13	1	5	2	17	2	5	2	23	3	5	2	32	4	5	2	56	5	5	2	540
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0	5	4	17	1	5	4	22	2	5	4	29	3	5	4	41	4	5	4	72	5	5	4	1600
0	5	5	19	1	5	5	24	2	5	5	32	3	5	5	45	4	5	5	81				